

# Intravitreal Resveratrol as Anti Apoptotic Agent Against Retinal Ganglion Cell Loss in Ischemic Reperfusion Injury

Amelia Shinta Prasetya<sup>1</sup>, Evelyn Komaratih<sup>1,\*</sup>, Wimbo Sasono<sup>1</sup>, Mercia Chrysanti<sup>1</sup>, Maria Debora Niken Larasati<sup>1</sup>, I Ketut Sudiana<sup>2</sup>

Amelia Shinta Prasetya<sup>1</sup>, Evelyn Komaratih<sup>1,\*</sup>, Wimbo Sasono<sup>1</sup>, Mercia Chrysanti<sup>1</sup>, Maria Debora Niken Larasati<sup>1</sup>, I Ketut Sudiana<sup>2</sup>

<sup>1</sup>Department of Ophthalmology, Faculty of Medicine, Universitas Airlangga/Dr. Soetomo General Hospital, Surabaya, INDONESIA.

<sup>2</sup>Departement of Anatomical Pathology, Faculty of Medicine, Universitas Airlangga, Surabaya, INDONESIA.

## Correspondence

Evelyn Komaratih

Department of Ophthalmology, Faculty of Medicine, Universitas Airlangga/Dr. Soetomo General Hospital, Surabaya, INDONESIA.

E-mail: evelyn.komaratih@fk.unair.ac.id

## History

- Submission Date: 29-09-2023;
- Review completed: 15-11-2023;
- Accepted Date: 27-11-2023.

DOI : 10.5530/pj.2023.15.219

## Article Available online

<http://www.phcogj.com/v15/i6>

## Copyright

© 2023 Phcogj.Com. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

## ABSTRACT

**Background:** Glaucoma is an optic neuropathy caused by the apoptosis of retinal ganglion cells and results in progressive retinal ganglion cell injury. A decrease in intraocular pressure (IOP) is a modifiable risk factor for slowing the progression of the disease, and can be accomplished through medication, laser therapy, or surgery. Even though the intraocular pressure has decreased and attained normal levels, the injury to the retinal ganglion cells continues in some cases. It is believed that neuroprotective administration has a positive effect on preventing the loss of retinal ganglion cells. **Methods:** Bax and Caspase-3 expression were measured involving 20 eyeballs of *Rattus Norvegicus* by immunohistochemistry examination. I-R injury was developed by increasing intraocular pressure (IOP) through the intracameral balanced salt solution (BSS) injection, then lowered after 60 minutes. Samples were divided into 4 groups: control, no further injection group, phosphate-buffered saline (PBS)-injected group and resveratrol-injected group. Each group was enucleated at days 7, 0, 7, and 7, respectively. Data with a non-normal distribution were examined using the Kruskal-Wallis test, and if the outcome was significant, the Mann-Whitney test. **Results:** The highest mean Bax and Caspase-3 expression was found in PBS injected and enucleated at day 7 group (G2),  $0.96 \pm 0.40$  and  $0.72 \pm 0.30$ , respectively. When compared to PBS injection, the expression of Bax and Caspase-3 was lower in the resveratrol-injected group. **Conclusion:** Bax and Caspase-3 expressions were lower in the intravitreal injection of Resveratrol in the dose of 100  $\mu\text{M}$  following the I-R injury group compared to the group without intravitreal Resveratrol injection. **Key words:** Ischemic-reperfusion injury, Glaucoma, Neuroprotective, Resveratrol, Apoptosis.

## INTRODUCTION

Glaucoma, a neurodegenerative condition, has specific features which are progressive retinal ganglion cell (RGC) and axon loss<sup>1</sup>. It is manifested in retinal nerve fiber layer thinning and optic nerve head (ONH) cupping, which functionally causes defects in the visual field starting from the peripheral and progressing to the central field<sup>2</sup>. Glaucoma affected more than 70 million people globally with approximately 10% with bilateral blindness<sup>3</sup>. Increase in intraocular pressure (IOP) is a well-known risk factor for glaucoma, due to its compressing mechanism which disrupts blood flow to the retina and optic nerve. This lead to tissue hypoxia and RGC death<sup>4,5</sup>. The IOP is related to ischemic-reperfusion (I-R) injury. It is started by ischemic injury which leads to hypoxia and hyponutrition. Metabolic acidosis starts to complicate the condition, following prolonged ischemia. On the other hand, reperfusion causes a resumption of blood flow, increase of local inflammation and reactive oxidative species (ROS), which causes secondary injury. Depending on how long the I-R injury has been present, the damaged cells may undergo apoptosis, autophagy, necrosis, and necroptosis<sup>6</sup>. The two pathways – intrinsic and extrinsic- are involve in RGC apoptosis. These routes connect to Caspase-3, a cysteine protease, and Bax, a Bcl-2 family<sup>6-10</sup>.

Glaucoma can be treated by medication, as the first line, followed by laser, and surgery. The aim is to reduce optic nerve damage progressivity and maintain patient's quality of

life. IOP reduction  $\geq 30\%$  is proven to prevent advanced optic nerve damage<sup>11</sup>. However, there are cases of glaucoma without increased IOP and continued disease progressivity in controlled IOP. Therefore, neuroprotective agents can be an alternative treatment<sup>1,12</sup>. One of the widely studied neuroprotective agents is resveratrol. Resveratrol, a natural phenol, is produced by plants in reaction to harm. Fruits like blueberries, mulberries, and raspberries contain resveratrol. Resveratrol had been studied in numerous eye diseases, for example, glaucoma, age-related macular degeneration (AMD), diabetic retinopathy, retinoblastoma, and retinopathy of prematurity (ROP)<sup>13,14</sup>.

Based on previously explained data, resveratrol plays role in glaucoma treatment, especially with I-R injury. In light of the quantity of Bax and Caspase expressions on RGC, the objective of this study is to comprehend the impact of resveratrol administration on RCG loss.

## MATERIALS AND METHODS

The *in vivo* experimental with post-test-only study was conducted in Universitas Airlangga from July 2022-January 2023. Mice models were used as the research subject, which were then divided into 4 groups through simple random sampling. The total research subjects were 20 eyeballs of *Rattus Norvegicus*. The groups were coded group G0, G1, G2, and G3. G0 was a control group with no treatment and enucleated at day 0. G1 was given I-R injury and enucleated at day 0 (IR0). G2 was given I-R injury, injected with intravitreal phosphate-

**Cite this article:** Prasetya AS, Komaratih E, Sasono W, Chrysanti M, Larasati MDN, Sudiana IK. Intravitreal Resveratrol as Anti Apoptotic Agent Against Retinal Ganglion Cell Loss in Ischemic Reperfusion Injury. *Pharmacogn J.* 2023;15(6): 1207-1212.

buffered saline (PBS), and enucleated at day 7 (IR7+PBS). G3 was done I-R injury, injected intravitreal Resveratrol, and enucleated at day 7 (IR7+RSV).

The research subject was healthy adult *Rattus Norvegicus*, weighing 250 - 300 grams, aged of 6-8 weeks. Those with any disease diagnosed by veterinarians or subjects with potentially transmitting disease during evaluation were excluded. The drop-out criteria were subject being sick or passing away during the experiment.

All animal procedures were carried out in accordance with The Animal Care and Use Committee of Universitas Airlangga of Veterinary Faculty (Ethical Clearance No: 2.KEH.138.10.2022). The research subjects were divided into four groups (G0-3). They were given intraperitoneal anesthesia ketamine hydrochloride 5% (80 mg/ kg) and xylazine hydrochloride (5 mg/kg). Afterward, they were given tetracaine hydrochloride 0.5% and disinfected with povidone iodine 5% on the ocular surface. G0 served as the control group. The IOP of G1-G3 was increased by injecting balanced salt solution (BSS) into the anterior chamber using canula of 30 G. The IOP was maintained at 110 mmHg for 60 minutes. G1 was enucleated at day 0. G2 was injected with PBS 2 µl intravitreal, then enucleated on day 7. G3 was injected with Resveratrol 100 µM in 2 µl intravitreal followed by enucleation on day 7. The retina layer from each group was taken to undergo immunohistochemistry examinations.

Bax and Caspase-3 expressions were examined using immunohistochemistry stain, the Bax primer monoclonal (Bioss Inc) and Caspase-3 primer monoclonal antibody (Bioss Inc), respectively. Both primers were diluted 1:100. Other than the primers, secondary antibodies were also used. It was examined under 400x microscope magnification in 625 micrometer power field.

### Data Analysis

Data were analyzed using SPSS 26.0. The distribution of the data was examined using Shapiro Wilk test. Oneway ANOVA was then used to evaluate normally distributed data. If the result was significant, it would be tested using posthoc Dunnet. Data with a non-normal distribution were analyzed using Kruskal-Wallis test, and if the outcome was significant, the Mann-Whitney test.

## RESULTS

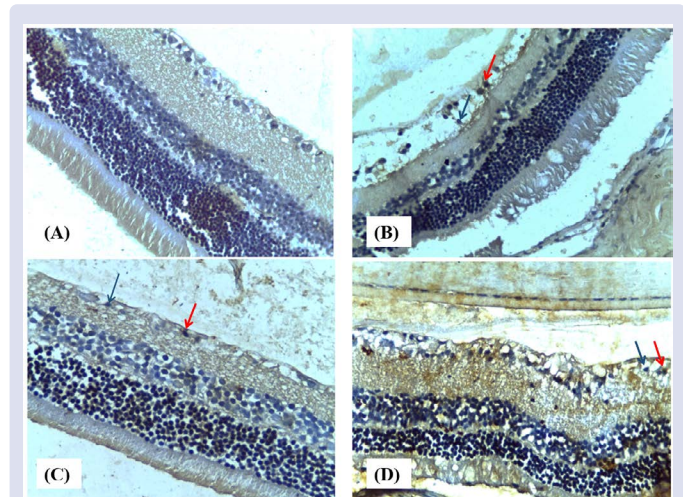
Immunohistochemistry examination showed positive RGC expressing Bax as brown at the cytoplasm. Figure 1 shows Bax-expressing cells in each experiment group.

Saphiro Wilk test resulted in the discovery of non-normal distribution. The values of the mean and standard deviation for the cell counts expressing Bax for each group are shown in Table 1. The highest Bax-expressing cell was found in G2. Analysis using Kruskal Wallis exhibits significant difference in median value in all groups with p-value = 0.003. Table 2 shows the analysis between groups using Mann Whitney test. In group comparison, G0 compared to G2, G0 to G3, G1 to G2, and G2 to G3 (p = 0.007, 0.008, and 0.033, respectively), Ba Bax expression in the group with resveratrol (G3) did not exhibit a significant difference compared to I-R injury (G1).

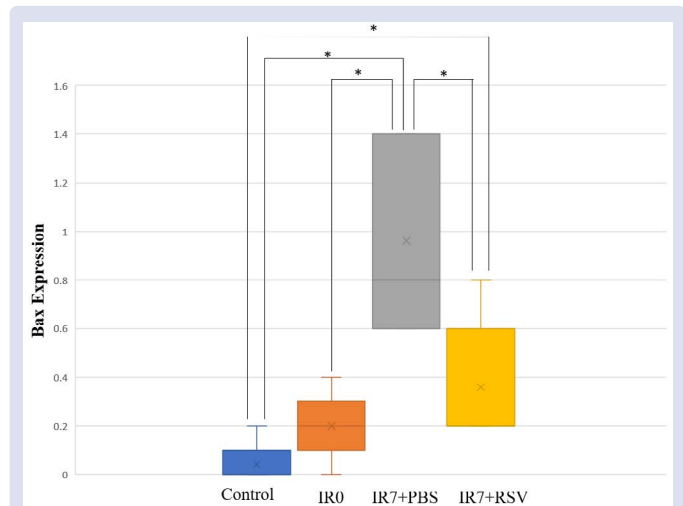
### Caspase-3 expression

Following immunohistochemistry staining using Caspase-3 primer and secondary antibody, Caspase-3 expressing cell was noted as brown at the cytoplasm. Figure 3 shows Bax-expressing cells in each group.

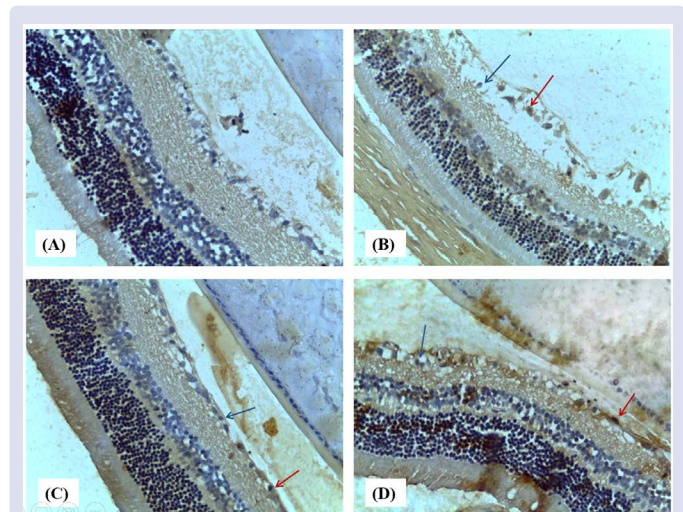
Normality test using Saphiro Wilk indicated non-normal distribution. Table 3 shows the value of mean ± standard deviation of cell number expressing Caspase-3 of each group. The highest Caspase-3-expressing cell was found in the IR7+PBS group or G2. Analysis using Kruskal



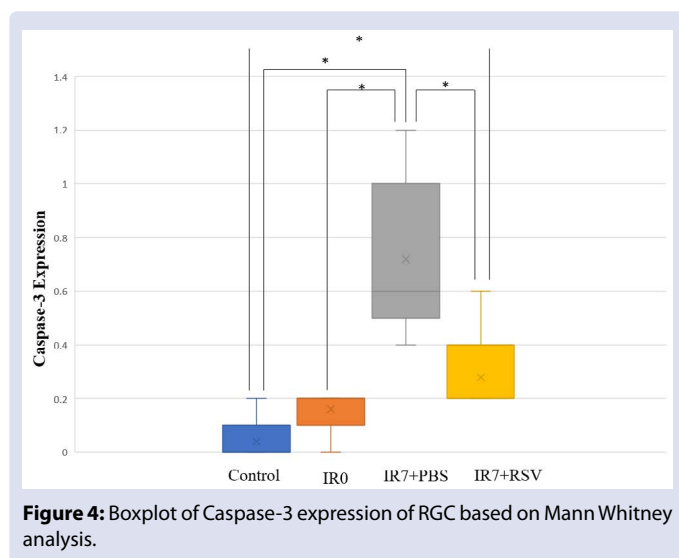
**Figure 1:** Bax expression (A) Control/ K0; (B) IR0; (C) IR7 + PBS; (D) IR7 + RSV. Bax-expressing cell (red arrow); non-Bax-expressing cell (blue arrow).



**Figure 2:** Boxplot of Bax expression of RGC based on Mann Whitney analysis.



**Figure 3:** Caspase-3 expression (A) Control/ K0; (B) IR0; (C) IR7 + PBS; (D) IR7 + RSV. Caspase-3-expressing cell (red arrow); non-Caspase-3-expressing cell (blue arrow).



**Figure 4:** Boxplot of Caspase-3 expression of RGC based on Mann Whitney analysis.

**Table 1: Distribution and basic characteristic retinal ganglion cell (RGC) expressing Bax.**

Group	N	Number of cells expressing Bax				P value
		Minimum	Maximum	Mean±SD	Median	
Control (G0)	5	0.00	0.20	0.04 ± 0.09	0.00	0.003*
IR0 (G1)	5	0.00	0.40	0.20 ± 0.14	0.20	
IR7 + PBS (G2)	5	0.60	1.40	0.96 ± 0.40	0.80	
IR7 + RSV (G3)	5	0.20	0.80	0.36 ± 0.26	0.20	

Statistically significant if \*p<0.05

**Notes:**

**Abbreviation:** IR0, I-R injury and enucleated at day 0; IR7+PBS, I-R injury given intravitreal PBS and enucleated at day 7; IR7+RSV, I-R injury given intravitreal Resveratrol and enucleated at day 7

**Table 2: Analysis of Bax expression in RGC between experiment groups.**

	Control (G0)	IR0 (G1)	IR7+PBS (G2)	IR7+RSV (G3)
Control (G0)		0.065	0.007*	0.014*
IR0 (G1)	0.065		0.008*	0.288
IR7+PBS (G2)	0.007*	0.008*		0.033*
IR7+RSV (G3)	0.014*	0.288	0.033*	

Statistically significant if \*p<0.05

**Notes:**

**Abbreviation:** IR0, I-R injury and enucleated at day 0; IR7+PBS, I-R injury given intravitreal PBS and enucleated at day 7; IR7+RSV, I-R injury given intravitreal Resveratrol and enucleated at day 7

**Table 3: Distribution and basic characteristic retinal ganglion cell (RGC) expressing Caspase-3.**

Group	N	Number of cells expressing Caspase-3				P value
		Minimum	Maximum	Mean±SD	Median	
Control (G0)	5	0.00	0.20	0.04 ± 0.09	0.00	0.002*
IR0 (G1)	5	0.00	0.20	0.16 ± 0.09	0.20	
IR7 + PBS (G2)	5	0.40	1.20	0.72 ± 0.30	0.60	
IR7 + RSV (G3)	5	0.20	0.60	0.28 ± 0.18	0.20	

Statistically significant if \*p<0.05

**Notes:**

**Abbreviation:** IR0, I-R injury and enucleated at day 0; IR7+PBS, I-R injury given intravitreal PBS and enucleated at day 7; IR7+RSV, I-R injury given intravitreal Resveratrol and enucleated at day 7

**Table 4: Analysis of Caspase-3 expression in RGC between experiment groups.**

	Control (G0)	IR0 (G1)	IR7+PBS (G2)	IR7+RSV (G3)
Control (G0)		0.072	0.007*	0.015*
IR0 (G1)	0.072		0.007*	0.180
IR7 + PBS (G2)	0.007*	0.007*		0.022*
IR7 + RSV (G3)	0.015*	0.180	0.022*	

Statistically significant if \*p<0.05

**Notes:**

**Abbreviation:** IR0, I-R injury and enucleated at day 0; IR7+PBS, I-R injury given intravitreal PBS and enucleated at day 7; IR7+RSV, I-R injury given intravitreal Resveratrol and enucleated at day 7

Wallis exhibits significant differences in median value with p = 0.002. Analysis was continued with Mann Whitney test and resulted in significant differences which can be seen in Table 4. Significant differences between G0 and G2 (p=0.007), G0 and G3 (p=0.015), G1 and G2 (p=0.007), as well as G2 and G3 (p=0.022) were discovered in group comparison. Between G1 and G3, however, no discernible differences were found. Figure 4 showing Caspase-3 expression in RGC is significantly higher at G2 compared to G0, G1, and G3. As of p-value = 0.180, there were no significant changes between G1 and G3.

**DISCUSSION**

Retina is a structure that converts light impulses into bioelectric signals in order to create visual information. Pathological alterations in the retinal ganglion cells occur during ischemia and hypoxia, resulting in retinal ganglion cell and optic nerve degeneration. Glaucoma is a progressive visual neuropathy defined by the loss of retinal ganglion cells<sup>15</sup>. An increase in IOP is the most common risk factor for glaucoma<sup>16</sup>. It limits oxygen supply and activates the apoptotic cascade of retinal ganglion cell death in situations of elevated IOP. Cell death is irreversible; retinal ganglion cells do not regenerate like central nervous system cells. Visual abnormalities and irreversible vision field loss will result from retinal ganglion cell death. Vision disturbances develop gradually, beginning in the periphery vision and progressing to the central vision, where they might impair patients' quality of life<sup>17-20</sup>.

Ischemia is defined as tissue hypoperfusion. This condition must be treated in advance by restoring perfusion to control cell damage and maintain organ function. However, the clinical outcome following reperfusion is not optimal and may induce secondary injury to the ischemic tissue. This phenomenon is called ischemic-reperfusion (I-R) injury<sup>21</sup>. The process of ischemia reperfusion injury is initiated by the occurrence of an ischemic state. The induction of anaerobic metabolism in tissues leads to the impairment of ion exchange mechanisms and a reduction in adenosine triphosphate (ATP) synthesis. The occurrence of ion exchange failure leads to cellular swelling and has a significant impact on the enzymatic activity inside the cytoplasm. Elevated intraocular pressure (IOP) leads to a decrease in oxygen delivery, triggering the initiation of cellular apoptosis. This process is initiated by heightened oxidative stress and the generation of reactive oxygen species (ROS)<sup>22</sup>, including superoxide anions (O<sub>2</sub><sup>-</sup>) and hydroxyl radicals (OH•), which then activate the cellular death pathway<sup>23,24</sup>. The intraocular pressure used in this investigation to cause I-R injury was 110 mmHg. As the pressure was kept for 60 minutes, hypoperfusion took place, followed by ischemia in the retinal layer. The pressure was lowered by cannula removal, hence creating tissue reperfusion condition. A previous study by Nurwasis et al., showed an increase in apoptotic retinal ganglion cell biomarkers such as SOD, HSP 80 RGC, and TNF-α RGC microglia, following increased IOP<sup>25</sup>. Studies implementing I-R injury had been done widely with different techniques. Luo et al used saline injection into the anterior chamber to increase the IOP<sup>26</sup>. Meanwhile Cao et al,

injected microbeads into the anterior chamber to increase IOP<sup>27</sup>. In this study, Ischemic-reperfusion injury achieved using saline injection to anterior chamber, the pressure increased to 110 mmHg which was higher than previous study and 60 minutes long to achieve higher number of injury. Saline injection to anterior chamber chosen of its ease to manage and also mimics increase IOP in glaucoma.

Several research have been conducted to investigate the various techniques for attaining the neuroprotective and therapeutic advantages of resveratrol in the treatment of glaucoma, either by peritoneal injection or oral administration. Previous studies have indicated that the bioavailability of orally administered resveratrol is below 1%. Prior to entering the systemic circulation, the concentration of free resveratrol undergoes a substantial reduction as a result of presystemic metabolism. In addition, it is important to take into account the blood-ocular barrier while giving resveratrol systemically for the treatment of eye diseases. The presence of resveratrol was seen only in the conjunctiva of 10 out of 35 eyes after the administration of oral trans-resveratrol. These previous studies prompted this study to use the intravitreal route for administering resveratrol dosage of 100  $\mu$ M to raise the intraocular resveratrol concentration<sup>17,26</sup>.

The result of this study showed that the highest mean of Bax-expressing cells was found in the group of I-R injury with PBS injection which was enucleated on the 7<sup>th</sup> day. The highest mean of Caspase-expressing cells was also found in the same group. Significant difference was found between (1) G0 and G2, (2) G0 and G3, and (3) G1 and G2. Increased expression of Bax indicates activation of apoptosis intrinsic pathway<sup>28</sup>. Increased expression of Caspase-3 indicates an ongoing apoptosis mechanism<sup>29</sup>. In the work by Luo et al., the elevation of Caspase-3 cleavage at day 3 was followed by an increase in Bax expression in the retina at 1 day following I-R injury<sup>26</sup>. The expression of Bax and Caspase-3 can be explained by apoptosis mechanism, the extrinsic and intrinsic pathways. Tumor necrosis factor/TNF- is one of the death ligands and receptors that activates the extrinsic route, commonly referred to as the death receptor pathway. These apoptosis-inducing complexes activate Caspase-8 protease to cleavage Caspase-3, of which will proteolysis the damaged cell. The pro-apoptotic Bcl-2 family is activated by the intrinsic pathway, also known as the mitochondrial pathway, which is initiated by hypoxia, radiation, or cellular toxins. These cascades will lead to apoptosis<sup>21</sup>.

Further findings show that Bax and Caspase-3 expression in the group with intravitreal resveratrol administration was lowered compared to PBS injected. The G1 and G3 groups did not differ significantly in any other ways. This indicates that resveratrol 100  $\mu$ M has a preventive effect in the occurrence of retinal ganglion cell death post I-R injury. This study is supported by Luo et al., whose findings indicate the preventive effect of Resveratrol in retinal ganglion cell apoptosis post-ischemia-reperfusion injury<sup>26</sup>. Similar method of administration was done by a previous study and showed that Resveratrol intravitreal administration has dose-dependent manner protective properties in retinal ganglion cells after I-R injury<sup>17</sup>. The mechanism is by blocking the proapoptotic pathway of Bax-cleavage and caspase-3<sup>26</sup>. Another postulated mechanism is that Resveratrol upregulates SIRT1 which inhibited RGC apoptosis, decreased Bax expression, and increased p-Akt expression<sup>17</sup>. It also inhibits the HIF-1 $\alpha$ /VEGF and p38/p53 pathway. Inhibiting these pathways helps to reduce RGC loss and retinal function impairment caused by retinal ischemia injury<sup>30</sup>.

Resveratrol, a polyphenol, exerts its effects via modulating a range of physiological processes, such as oxidative stress, cell proliferation, apoptosis, inflammation, metastasis, and angiogenesis<sup>31</sup>. Resveratrol has several benefits, including but not limited to anti-aging, anti-cancer, anti-diabetic, neuroprotective, cardioprotective, wound healing, and therapeutic potential for depressive symptoms<sup>32-34</sup>. It works by activating Sirtuin 1 or SIRT1. SIRT1 activation is known to be

RGC neuroprotective by suppressing (I-R) injury apoptotic pathway. RGC survival rates and SIRT1 expression decline with I-R damage duration<sup>17</sup>.

Based on study results, it has been demonstrated that resveratrol exhibits the ability to counteract the phenomenon of apoptosis in SGR. Resveratrol functions by the activation of SIRT1, a member of the nicotinamide adenine dinucleotide (NAD<sup>+</sup>)-dependent histone deacetylase family, which is recognized for its role in the modulation of cellular lifespan<sup>35</sup>. The enhancement of the antioxidant response to mitochondrial failure is achieved by the upregulation of SIRT1, resulting in a reduction in the expression of Bax, a known inducer of apoptosis. This reduction in Bax expression leads to the inhibition of its release of Cytochrome C, therefore preventing the activation of caspase-3, a crucial step in the execution of apoptosis<sup>36</sup>.

This study has a potential limitation. The route of Resveratrol administration in this study was intravitreal, an invasive method with potential side effects such as increased intraocular pressure, vitreous hemorrhage, and retinal ablation.

Further research should be conducted using other cell death markers, for example, Bcl-2, HSP, and Caspase-9. Other routes of resveratrol administration should be tried, particularly non-invasive methods such as eye drops or nanoparticles. These other routes should be able to maintain resveratrol bioavailability. Further research can be conducted on bigger experimental animals or even on humans.

## CONCLUSION

This experimental study on *Rattus Norvegicus* was aimed at finding the effect of resveratrol in anti apoptic or cell death pathway by evaluating Bax and Caspase-3 expression in retinal ganglion cell loss following I-R injury. Together, it was demonstrated that the expressions of Bax and Caspase-3 were reduced in the group that received an intravitreal injection of resveratrol after suffering an I-R injury as compared to the group that did not receive an intravitreal resveratrol injection. This study also found that retinal ganglion cell death following I-R injury can be prevented by using Resveratrol at the dose of 100  $\mu$ M. In conclusion, this experimental study indicates the utility of Resveratrol in I-R injury, which can be further applied in glaucoma medication.

## ACKNOWLEDGEMENT

Dr. Nurwasis dr. SpM(K), Dr. Yulia Primitasari dr. Sp.M(K), and Dr. Titiek Ernawati dr., SpM(K) for guidance in glaucoma model and research method. Apt. Christawan Ardianto, S.Farm., M.Sc., Ph.D and Faculty of Pharmacy for the assistance, materials, and facilities for this research. Dr. Rachmah Indawati, SKM. MKM (Faculty of Public Health – Universitas Airlangga) for statistical and methodology advice. Djoko Legowo, Mke., drh (Faculty of Veterinary Medicine – Universitas Airlangga) for ethical clearance, preparation of animal and pathological anatomy prepare. The Indonesian Ministry of Research and Higher Education provided funding for this study.

## DISCLOSURE

The author reports no conflicts of interest in this work.

## AUTHOR CONTRIBUTION

All authors contributed equally to this research and publication of this manuscript.

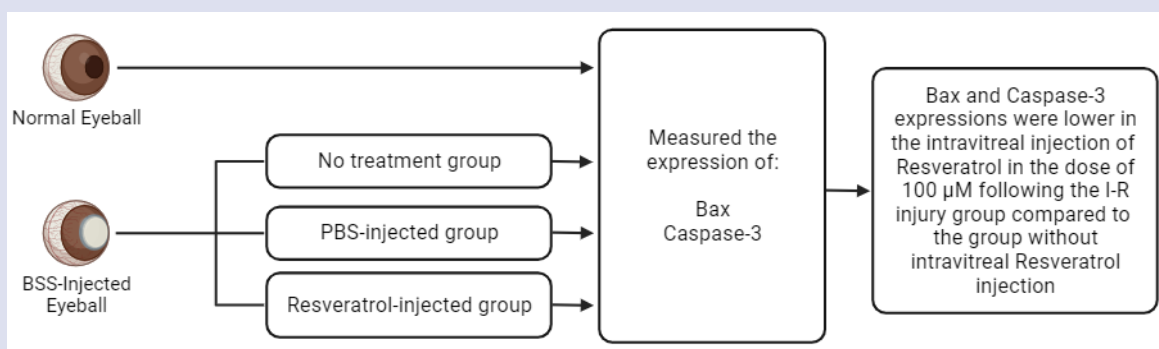
## REFERENCES

1. Russo R, Varano GP, Adornetto A, Nucci C, Corasaniti MT, Bagetta G, et al. Retinal ganglion cell death in glaucoma: Exploring the role of neuroinflammation. *Eur J Pharmacol.* 2016 Sep;787:134–42. Available from: <https://doi.org/10.1016/j.ejphar.2016.03.064>

2. Johnson T V., Tomarev SI. Animal Models of Glaucoma. In 2016. p. 31–50. Available from: [https://doi.org/10.1007/978-3-319-19434-9\\_3](https://doi.org/10.1007/978-3-319-19434-9_3)
3. Kementerian Kesehatan RI. Situasi dan Analisis GLAUKOMA. InfoDATIN Pusat Data dan Informasi Kementerian Kesehatan RI. 2015. p. 1–6.
4. Rumelt S. Glaucoma - Basic and Clinical Concept. Rumelt S, editor. InTech; 2011. Available from: <https://doi.org/10.5772/792>
5. Ung L, Pattamatta U, Carnt N, Wilkinson-Berka JL, Liew G, White AJR. Oxidative stress and reactive oxygen species: a review of their role in ocular disease. *Clin Sci*. 2017 Dec 15;131(24):2865–83. Available from: <https://doi.org/10.1042/cs20171246>
6. Wu MY, Yang GT, Liao WT, Tsai APY, Cheng YL, Cheng PW, et al. Current Mechanistic Concepts in Ischemia and Reperfusion Injury. *Cellular Physiology and Biochemistry*. 2018;46(4):1650–67. Available from: <https://doi.org/10.1159/000489241>
7. Aslan M, Dogan S, Kucuksayan E. Oxidative stress and potential applications of free radical scavengers in glaucoma. *Redox Report*. 2013 Mar 19;18(2):76–87. Available from: <https://doi.org/10.1179/1351000212y.0000000033>
8. Pawlowski J, Kraft AS. Bax-induced apoptotic cell death. *Proceedings of the National Academy of Sciences*. 2000 Jan 18;97(2):529–31. Available from: <https://doi.org/10.1073/pnas.97.2.529>
9. Kiraz Y, Adan A, Kartal Yandim M, Baran Y. Major apoptotic mechanisms and genes involved in apoptosis. *Tumor Biology*. 2016 Jul 9;37(7):8471–86. Available from: <https://doi.org/10.1007/s13277-016-5035-9>
10. Artini IGA, Astuti KW, Trapika IGMGSC, Indrayani AW. The bcl-2 and caspase-3 expression after purple sweet potato treatment on isoniazid and rifampicin-induced liver injury. *Bali Medical Journal*. 2023;12(2):1204–10. Available from: <https://doi.org/10.15562/bmj.v12i2.437>
11. Wills Eye Hospital. The Wills Eye Manual: Office and Emergency Room Diagnosis and Treatment of Eye Disease. 7th ed. Bagheri N, Wajda BN, Calvo C, editors. Philadelphia: Wolters Kluwer; 2017.
12. Pirhan D, Yüksel N, Emre E, Cengiz A, Kürşat Yıldız D. Riluzole and Resveratrol-Induced Delay of Retinal Ganglion Cell Death in an Experimental Model of Glaucoma. *Curr Eye Res*. 2016 Jan 2;41(1):59–69. Available from: <https://doi.org/10.3109/02713683.2015.1004719>
13. Abu-Amero K, Kondkar A, Chalam K. Resveratrol and Ophthalmic Diseases. *Nutrients*. 2016 Apr 5;8(4):200. Available from: <https://doi.org/10.3390/nu8040200>
14. Delmas D, Cornebise C, Courtaut F, Xiao J, Aires V. New Highlights of Resveratrol: A Review of Properties against Ocular Diseases. *Int J Mol Sci*. 2021 Jan 28;22(3):1295. Available from: <https://doi.org/10.3390/ijms22031295>
15. Maharani M, Dewi PK, Prihatningtias R, Wildan A, Nugroho T, Limijadi EKS, et al. Aqueous Humour Malondialdehyde Level as Oxidative Stress Marker In Types Of Glaucoma. *Bali Medical Journal*. 2022 Mar 14;11(1):103–5. Available from: <http://dx.doi.org/10.15562/bmj.v11i1.2599>
16. Elsa Gustianty, Novaqua Yandi, R. Maula Rifada, Andika Prahasta. Successful rate of glaucoma surgery in uveitis glaucoma. *Bali Medical Journal*. 2022 Jul 19;11(2):614–8. Available from: <http://dx.doi.org/10.15562/bmj.v11i2.3568>
17. Luo J, He T, Yang J, Yang N, Li Z, Xing Y. SIRT1 is required for the neuroprotection of resveratrol on retinal ganglion cells after retinal ischemia-reperfusion injury in mice. *Graefes's Archive for Clinical and Experimental Ophthalmology*. 2020 Feb 3;258(2):335–44. Available from: <https://doi.org/10.1007/s00417-019-04580-z>
18. Corral-Domenge C, de la Villa P, Mansilla A, Germain F. Tools and Biomarkers for the Study of Retinal Ganglion Cell Degeneration. *Int J Mol Sci*. 2022 Apr 13;23(8):4287. Available from: <https://doi.org/10.3390/ijms23084287>
19. Almasieh M, Wilson AM, Morquette B, Cueva Vargas JL, Di Polo A. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res*. 2012 Mar;31(2):152–81. Available from: <https://doi.org/10.1016/j.preteyeres.2011.11.002>
20. Xia J, Yang X, Chen W. Resveratrol protects the retina from I/R injury by inhibiting RGCS apoptosis, glial activation and expression of inflammatory factors. *Tropical Journal of Pharmaceutical Research*. 2020 Nov 13;19(6):1221–6. Available from: <https://doi.org/10.4314/tjpr.v19i6.16>
21. Wu W, He J, Shao X. Incidence and mortality trend of congenital heart disease at the global, regional, and national level, 1990–2017. *Medicine*. 2020 Jun 5;99(23):e20593. Available from: <https://doi.org/10.1097/md.00000000000020593>
22. Dizaj SM, Sharifi S, Shahi S, Montazersaheb S, Salatin S, Ahmadian E, et al. The most important consideration in clinical usage of curcumin. *Journal of Medicinal and Pharmaceutical Chemistry Research*. 2022;4(2):124–36. Available from: <https://doi.org/10.22034/ecc.2022.319668.1278>
23. Al-Hameed NM, Al-Ani AW. Assessment of systemic oxidative stress and antioxidants in Iraqi women with newly diagnosed and tamoxifen-treated breast cancer. *Journal of Medicinal and Pharmaceutical Chemistry Research*. 2023;5(9):204–15. Available from: <https://doi.org/10.22034/ecc.2023.365482.1538>
24. Sudiana IK. Hantaran Sinyal pada Proses Inflamasi. Surabaya: Airlangga University Press; 2017.
25. Nurwasis, Suhendro G, Sudiana IK. The Role of SOD, Catalase, HSP-27, HSP-70, and TNF- $\alpha$  Expression in Apoptosis of Retinal Ganglion Cells After Intra Ocular Pressure Increase on Rattus Norvegicus. *Indian J Public Health Res Dev*. 2019;10(7):1174. Available from: <https://doi.org/10.5958/0976-5506.2019.01743.1>
26. Luo H, Zhuang J, Hu P, Ye W, Chen S, Pang Y, et al. Resveratrol Delays Retinal Ganglion Cell Loss and Attenuates Gliosis-Related Inflammation From Ischemia-Reperfusion Injury. *Investigative Ophthalmology & Visual Science*. 2018 Aug 1;59(10):3879. Available from: <https://doi.org/10.1167/iovs.18-23806>
27. Cao K, Ishida T, Fang Y, Shinohara K, Li X, Nagaoka N, et al. Protection of the Retinal Ganglion Cells: Intravitreal Injection of Resveratrol in Mouse Model of Ocular Hypertension. *Investigative Ophthalmology & Visual Science*. 2020 Mar 16;61(3):13. Available from: <https://doi.org/10.1167/iovs.61.3.13>
28. Alimoradzadeh R, Moosavi N, Karimkoshteh A, Sadeghi Z, Milanifard M, Ismaili A. Investigation of the Chemistry of Metformin by Targeting the Nrf2 Signaling Pathway (A response Surface Methodology Approach). *Chemical Methodologies*. 2022;6(3):166–73. Available from: <https://doi.org/10.22034/chemm.2022.315764.1395>
29. Yadav S, Sharma M, Ganesh N, Srivastava S, Srivastava MM. Bioactive principle loaded gold nanoparticles as potent anti-melanoma agent: green synthesis, characterization, and in vitro bioefficacy. *Asian Journal of Green Chemistry*. 2019 Jul 20;3(4):492–507. Available from: [10.33945/SAMI/AJGC.2019.4.6](https://doi.org/10.33945/SAMI/AJGC.2019.4.6)
30. Ji K, Li Z, Lei Y, Xu W, Ouyang L, He T, et al. Resveratrol attenuates retinal ganglion cell loss in a mouse model of retinal ischemia reperfusion injury via multiple pathways. *Exp Eye Res*. 2021 Aug;209:108683. Available from: <https://doi.org/10.1016/j.exer.2021.108683>
31. Kasim SM, Abdulaziz NT, Jasim MH, Mustafa YF. Resveratrol in cancer chemotherapy: Is it a preventer, protector, or fighter? *Journal of Medicinal and Pharmaceutical Chemistry Research*. 2023;5(7):576–87. Available from: <https://doi.org/10.22034/ecc.2023.379480.1586>
32. Prasetya AS, Komaratih E, Sasono W, Chrysanti M, Larasati MDN, Sudiana IK. The role of Resveratrol as a neuroprotective agent in the prevention of retinal ganglion cell loss in ischemic reperfusion injury animal model: a literature review. *Bali Medical Journal*. 2023;12(2):1796–801. Available from: <https://doi.org/10.15562/bmj.v12i1.4449>

33. Ardianto C, Budiati AS, Sumartha INB, Nurrahmi N, Rahmadi M, Khotib J. Resveratrol ameliorates physical and psychological stress-induced depressive-like behavior. *J Basic Clin Physiol Pharmacol*. 2021 Jun 25;32(4):335–40. Available from: <https://doi.org/10.1515/jbcpp-2020-0437>
34. Prakoeswa CRS, Rindiastuti Y, Wirohadidjojo YW, Komaratih E, Nurwasis, Dinaryati A, et al. Resveratrol promotes secretion of wound healing related growth factors of mesenchymal stem cells originated from adult and fetal tissues. *Artif Cells Nanomed Biotechnol*. 2020 Jan 1;48(1):1159–66. Available from: <https://doi.org/10.1080/21691401.2020.1817057>
35. Setianingsih H, Soetjipto S, Sudiana IK, Suryokusumo MG. Hyperbaric oxygen effects towards SIRT1 level in Sprague dawley with endothelial dysfunction by high-cholesterol diet. *Bali Medical Journal*. 2018 Aug 1;7(2). Available from: <http://dx.doi.org/10.15562/bmj.v7i2.1154>
36. Balaiya S, Abu-Amero KK, Kondkar AA, Chalam K V. Sirtuins Expression and Their Role in Retinal Diseases. *Oxid Med Cell Longev*. 2017;2017:1–11. Available from: <https://doi.org/10.1155/2017/3187594>

## GRAPHICAL ABSTRACT



**Cite this article:** Prasetya AS, Komaratih E, Sasono W, Chrysanti M, Larasati MDN, Sudiana IK. Intravitreal Resveratrol as Anti Apoptotic Agent Against Retinal Ganglion Cell Loss in Ischemic Reperfusion Injury. *Pharmacogn J*. 2023;15(6): 1207-1212.