Hypoglycemic Activity of Leaf Extracts from *Tiliacora triandra* in Normal and Streptozotocin-Induced Diabetic Rats

Teeraporn Katisart¹, Surapong Rattana²

INTRODUCTION

Diabetes mellitus (DM) is a metabolic disease characterized by hyperglycemia. It is caused by insulin secretion deficiency or disorder of insulin function. The long term diabetes leads to the complications in various body systems such as diabetic retinopathy, nephropathy and neuropathy.¹ It is estimated that in the year 2025, there will be 300 million patients suffered from diabetes. This amount is rising in the developing countries including Thailand.² The government has taken into account to spend budget for production and import of chemically synthetic drugs such as insulin, glibenclamide, metformin and acarbose. The interesting alternative is to use traditional medicine for treatment of diabetes.

*Tiliacora triandra* is a medicinal plant belonging to the family Menispermaceae. It is a common indigenous plants found in south-east Asia including Thailand.³ The leaves of *Tiliacora triandra* has been used as an ingredient in young bamboo soup because it could reduce the toxicity of bamboo shoot. It has been used in Thai traditional medicine for the treatment of fever.⁴ Pharmacological study of leaf extracts revealed that it showed high anti-oxidant capacity in accordance with high flavonoid and phenolic compound contents.⁵,⁶ These findings corresponded to the study by Singthong et al. who found that water leaf extract of *Tiliacora triandra* provided phenolic compound and showed high antioxidant activities.⁷ Kaewpiboon et al. found that three fatty acid from leaf extract of *Tiliacora triandra* enhanced P-glycoprotein function in multidrug-resistant A549RT-eto cell line.⁸ Aerial part extract of *Tiliacora triandra* is effective in treating brain dysfunction induced by alcohol and possess anti-tumor and anti-cancer activity.⁹–¹¹ Alkaloids from *Tiliacora triandra* also exhibit antimycobacterial activity against multidrug-resistant isolates of *Mycobacterium tuberculosis*.¹²–¹³ However, the root extract of this plant contained two pure alkaloid compounds called tiliacorinine and tilicotine with anti-malarial activity.¹³,¹⁴ In addition, the root extracts also exhibit anti-pyretic activity.¹³,¹⁶ For toxicity study, it was found that the water extract from this plant does not cause acute or subchronic toxicities in either male or female rats.¹⁷ In addition, leaf extract from *Tiliacora triandra* showed alpha-glucosidase inhibitory activity suggesting the decrease of glucose absorption in small intestine. This finding suggested the potential of this plant in prevention of hyperglycemia.¹⁸ However, there is no report on anti-diabetic activity of this plant extracts in animal models. Therefore, the objectives of this study are to study the hypoglycemic effect of ethanol leaf extracts from *Tiliacora triandra* and to study the
possible mechanism of action of these extracts in lowering blood glucose level in animal model of diabetes.

MATERIALS AND METHODS

Preparation of plant extracts

*Tiliacora triandra* was cultivated in Mahasarakham province, Northeastern part of Thailand. The specimen was deposited at Department of Biology, Faculty of Science, Mahasarakham University, Thailand (Code: MSUSC-BI-TK1). The leaves were collected and dried in the hot air oven at 50 °C for 48 h. The leaves were ground as powder (1,000 g) and then macerated with 95% ethanol (4,000 ml) for 7 days. The solvent was removed using a rotary evaporator. The extracts were then dried using a freeze dryer to get a powder. The 95% ethanolic leaf extracts (TTE) were kept at -20 °C until use.

Animals

Male albino wistar rats were purchased from National Laboratory Animal Centre, Mahidol University, Thailand. The rats with body weight of 180-200 g were used. All rats were kept in separate cages. One control and one diabetic rat were kept in the same cage. Feed and water were provided daily to the rats for up to 8 weeks. All groups were kept in a temperature-controlled room (22 ± 2 °C), artificially lit from 6.00 to 18.00 hours daily. The initial weights and blood glucose levels of the rats were recorded weekly. The experimental protocol was approved by Mahasarakham University ethics committee, Mahasarakham, Thailand (License No. 0015/2015).

Induction of diabetes in rats

The rats were fasted for 8-12 h. The initial blood glucose level of rats from the tail vein was then measured using an Accu-Check active testing kit. Streptozotocin (STZ) was freshly dissolved in 20 mM citrate buffer (pH 4.5) STZ was injected once and intraperitoneally at dose of 65 mg/kg body weight to the rats. Then, 20 mM citrate buffer (pH 4.5) at an equal volume to the diabetic group was injected to the control rats. To avoid the initial hypoglycemia, 2% sucrose was added to drinking water for the streptozotocin-induced diabetic rats for 48 hours. Their blood glucose levels were measured immediately after one week of injection. The blood glucose level in control rats should be approximately 80 mg/dL. For diabetic groups, the blood glucose level of 126 mg/dL or more is confirmed as diabetic.

Experimental designs

The rats were divided into five experimental groups with six animals in each: group 1 was normal control rats treated orally with 0.5% tween 80; group 2 was normal rats treated with leaf extracts (300 mg/kg b.w.); group 3 was diabetic control rats treated orally with 0.5% tween 80; group 4 was diabetic rats treated orally with glibenclamide (0.25 mg/kg b.w.) and group 5 was diabetic rats treated with leaf extracts (300 mg/kg b.w. TTE and glibenclamide were suspended in 0.5% tween 80 and administered orally using orogastric tube daily for 8 weeks.

Effect of TTE on body weight and fasting plasma glucose

The normal and STZ-induced diabetic rats were fasted for 8-12 hours. Then blood samples were collected from the tail vein of the rats by using Accu-check Advantage II (Roche, Germany). FPG were measured weekly in each experimental groups.

Effect of TTE on serum insulin

Eight weeks after experiments, all the rats were fasted for 8-12 hours. They were sacrificed by cervical dislocation technique. Then, blood samples were drawn from the rat’s heart. The blood samples were centrifuged at 3500 rpm for 20 min to get serum. The serum samples were analyzed for insulin content by using immune radio assay kit (MP Bio chemicals-Orangeburg, USA).

Histological studies

The procedure of histological studies of pancreas was adapted according to the study by Zhang et al. (2010). After blood collection, all rats were sacrificed. The pancreatic tissues were removed. The tissues were then fixed in 10% buffered formalin after washing with normal saline. The tissues were processed for embedding in paraffin wax by routine protocols and 5-μm-thick sections were then cut by microtome. The tissues were stained with haematoxylin-eosin using a routine protocol and examined using photomicroscope.

Statistical analysis

All data were expressed as mean ± standard error of mean (SEM). Statistical analysis was carried out using F-test (One-way ANOVA) followed by Scheffe’s test. The criterion for statistical significance was at a p-value less than 0.05.

RESULTS

Effect of extracts on body weight

The body weight of rats (as shown in Table 1) were measured weekly along 8 weeks of the experiments. The results showed that normal and diabetic rats treated with 300 mg/kg b.w. extracts had an increased body weight week by week in comparison with the normal controls. However, the diabetic control rats had the decreased body weight compared to the normal controls.

Effect of extracts on fasting plasma glucose

The fasting plasma glucose (FPG) of rats (as shown in Table 2) were also measured weekly along 8 weeks of the experiments. The results showed that normal and diabetic rats treated with 300 mg/kg b.w. extracts had the decreased fasting plasma glucose week by week in comparison with the normal controls. However, the diabetic control rats had an increased fasting plasma glucose compared to the normal controls. In this study, possible mechanism of action of these extracts in lowering blood glucose level in animal model of diabetes.

### Table 1: The effect of ethanolic extract of *Tiliacora triandra* on body weight in normal and streptozotocin-induced diabetic rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Initial BW</th>
<th>Week 1</th>
<th>Week 2</th>
<th>Week 3</th>
<th>Week 4</th>
<th>Week 5</th>
<th>Week 6</th>
<th>Week 7</th>
<th>Week 8</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cont.</td>
<td>302.00±3.65</td>
<td>5.50±0.71</td>
<td>19.13±0.95</td>
<td>28.03±1.50</td>
<td>35.81±1.51</td>
<td>44.45±1.59</td>
<td>48.97±1.49</td>
<td>52.67±1.78</td>
<td>57.56±1.55</td>
</tr>
<tr>
<td>Cont.+TTE</td>
<td>303.00±8.65</td>
<td>3.31±0.58</td>
<td>8.10±1.35</td>
<td>17.35±1.74</td>
<td>26.51±2.39</td>
<td>30.75±1.13</td>
<td>40.74±2.83</td>
<td>44.13±2.93</td>
<td>48.63±3.10</td>
</tr>
<tr>
<td>DM</td>
<td>212.00±11.74</td>
<td>3.86±0.25</td>
<td>12.78±3.63</td>
<td>13.66±3.15</td>
<td>15.24±3.21</td>
<td>16.93±3.34</td>
<td>0.00±0.00</td>
<td>17.13±2.74</td>
<td>17.34±2.93</td>
</tr>
<tr>
<td>DM+GB</td>
<td>247.00±27.66</td>
<td>3.06±0.48</td>
<td>13.84±2.10</td>
<td>16.88±1.60</td>
<td>21.39±3.62</td>
<td>24.42±4.02</td>
<td>27.64±5.50</td>
<td>30.35±5.87</td>
<td>35.52±5.22</td>
</tr>
<tr>
<td>DM+TTE</td>
<td>207.00±83.28</td>
<td>4.78±1.80</td>
<td>12.57±1.80</td>
<td>16.14±2.40</td>
<td>0.00±0.00</td>
<td>0.00±0.00</td>
<td>21.81±5.04</td>
<td>25.81±4.40</td>
<td>31.55±6.40</td>
</tr>
</tbody>
</table>
The diabetic rats treated with leaf extracts had the improved histological appearances of pancreas.

**DISCUSSION**

Hyperglycemia is a remarkable characteristic among diabetic patients. It usually alters the balance of free radicals and antioxidant levels. This may lead to the damage of pancreatic beta cells and induction of insulin resistance. STZ has the cytotoxic effect on pancreatic beta cells. It may be mediated by the inhibition of free radical scavenging enzymes which is related to the production of superoxide radical. This may be evident by a decrease of pancreatic islets size, degranulation of cells, vacuolation and invasion of connective tissues in those of the diabetic control group. The possible mechanism of blood glucose reduction produced by plant extracts could be explained by different mechanisms. Some plants may contain insulin-like substances. The others may increase β cells in the pancreas by activating the regeneration of these cells. The fiber of plants may also interfere with carbohydrate absorption and therefore affect blood glucose.

A medicinal plant in the family Menispermaceae such as *Tinospora crispa* also exhibit hypoglycemic effect with insulin enhancing activity. However, hypoglycemic effect of *Tinospora cordifolia* may be through the glycogen storage in the liver or decreasing the glucose release from the liver. In the present study, it is clear that leaf extracts of *Tiliacora triandra* reduce blood glucose level in diabetic rats over eight weeks of the treatment. The extracts also increase body weight of all treatment groups. Treatment of the extracts causes the regeneration and histological improvement of pancreatic islets. This is confirmed by the elevation of serum insulin level in diabetic rats treated with the extracts. Therefore, it is assumed that the active ingredient of the extracts may act as insulin.

**Figure 1:** Histological illustration of rat pancreas stained with hematoxylin-eosin. (a) normal rats treated with 0.5% tween 80. (b) normal rats treated with TTE 300 mg/kg. (c) diabetic rats treated with 0.5% tween 80. (d) diabetic rats treated with TTE 300 mg/kg. (e) diabetic rats treated with glibenclamide 0.5 mg/kg.

**Table 2:** The effect of ethanolic extract of *Tiliacora triandra* on fasting blood glucose in normal and streptozotocin-induced diabetic rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Initial FBG (mg/dL)</th>
<th>Week 1</th>
<th>Week 2</th>
<th>Week 3</th>
<th>Week 4</th>
<th>Week 5</th>
<th>Week 6</th>
<th>Week 7</th>
<th>Week 8</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cont.</td>
<td>84.00±1.53</td>
<td>0.00±0.00⁶</td>
<td>0.00±0.00⁶</td>
<td>0.00±0.00⁶</td>
<td>0.00±0.00⁶</td>
<td>0.00±0.00⁶</td>
<td>0.00±0.00⁶</td>
<td>0.00±0.00⁶</td>
<td></td>
</tr>
<tr>
<td>DM</td>
<td>20.88±11.89</td>
<td>20.88±11.89</td>
<td>11.05±6.29⁶</td>
<td>11.05±6.29⁶</td>
<td>11.05±6.29⁶</td>
<td>11.05±6.29⁶</td>
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<td>11.05±6.29⁶</td>
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</tr>
<tr>
<td>DM+TTE</td>
<td>482.00±55.67</td>
<td>22.77±4.76⁶</td>
<td>22.77±4.76⁶</td>
<td>22.77±4.76⁶</td>
<td>22.77±4.76⁶</td>
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<td>22.77±4.76⁶</td>
<td></td>
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</tbody>
</table>

**Table 3:** The effect of ethanolic extract of *Tiliacora triandra* on serum insulin levels in normal and streptozotocin-induced diabetic rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Insulin level (IU/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cont.</td>
<td>22.77±0.23⁶</td>
</tr>
<tr>
<td>Cont.+TTE</td>
<td>23.02±0.64⁶</td>
</tr>
<tr>
<td>DM</td>
<td>10.63±0.37⁶</td>
</tr>
<tr>
<td>DM+TTE</td>
<td>21.63±1.39⁶</td>
</tr>
<tr>
<td>DM+GB</td>
<td>21.22±1.87⁶</td>
</tr>
</tbody>
</table>

The possible mechanism of blood glucose reduction produced by plant extracts could be explained by different mechanisms. Some plants may contain insulin-like substances. The others may increase β cells in the pancreas by activating the regeneration of these cells. The fiber of plants may also interfere with carbohydrate absorption and therefore affect blood glucose.
sensitizer which stimulates insulin secretion from the pancreas which in turn reduce or control the blood glucose levels.

CONCLUSION

The findings from this research can be concluded that leaf extracts from *Tiliacora triandra* exhibits the hypoglycemic potential by stimulating insulin secretion from the pancreas in STZ-induced diabetic rats without the significant hypoglycemic effect and body weight alterations in normal rats.

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CONFLICT OF INTEREST

Authors declare no conflict of interest.

ABBREVIATIONS USED

FBG: fasting blood glucose; BW: body weight; TTE: *Tiliacora triandra* leaf extract; STZ: Streptozotocin; GB: Glibenclamide; Cont.: control; DM: diabetes mellitus; IL: Islets of Langerhans.

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AUTHOR PROFILE

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